

Detection of Toxin Genes in E. Coli isolate PE74 by PCR analysis

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Abstract: — E.coli isolate pe74 culture which showed positive for cytotoxic effect was selected for PCR analysis. On genomic DNA isolation, isolate pe74 showed prominent band on agarose gel electrophoresis. Isolate pe74 was analyzed for the presence of *stx*, *stx2f*, *stx2*, *stx2c*, *stx2e*, *eaeA*, *stx2dandhlyA* using different primers. On PCR amplification isolate pe74 showed no amplified product for *Stx1* however only primer dimers were observed in sample. PCR analysis of isolate pe74 showed positive to *stx2f* with a band close to 150 bp and a band between 100 and 200 bp for *stx2c*.

Key words: E. coli, PCR analysis, toxin production

I. INTRODUCTION

Escherichia coli cells are rod shaped, gram negative, facultative anaerobic bacteria found in lower intestine of human and animals that grows at pH 7, temperature 37°C. Many strains of E. coli produce a variety of toxins, including Shiga-like toxins (*stx*). These strains are often referred to as Verotoxin-producing E. coli. Such toxigenic isolates have been identified to cause serious human gastrointestinal disease, often with severe complicating problems that include bloody diarrhea, life threatening condition hemolytic-uremic syndrome (Xiaodong et al., 2010). In the present study eight genes of E. coli are targeted for PCR confirmation, namely *stx*, *stx2f*, *stx2*, *stx2c*, *stx2e*, *eaeA*, *stx2dandhlyA*. The first major outbreak of STEC O157:H7 infection was first reported in the United States in 1982 and was linked to eating undercooked ground beef from fast-food restaurant chain (Riley et al., 1983). Based on the results obtained, isolate pe74 which showed positive to *stx2f* and *stx2c* was further subjected to purification method. Hence the study of toxins gene in E. coli isolate was carried out.

II. MATERIALS AND METHODS

2.1E. coli CULTURE

E. coli isolate pe74 was sub-cultured on Eosin Methylene Blue agar media. Colonies with green metallic sheen on EMB were selected for present study. E. coli culture pe74 was inoculated into 250 ml nutrient broth and was

incubated over night at room temperature with gentle shaking was subjected to Genomic DNA procedure.

2.2 GENOMIC DNA EXTRACTION

The pellet from 1.5 ml of overnight culture was resuspended in 500 µl of lysis buffer and incubated at 37°C for 1 hour to lyse the cells. Genomic DNA was then extracted by Chloroform. DNA from the aqueous phase was precipitated with isopropanol, washed with 70% Ethanol and air dried. The DNA pellet was dissolved in 50 µl of TE buffer. 1 µl of genomic DNA was used to analyze on 0.8% Agarose gel electrophoresis (Sambrook and Russell 1989).

2.3 POLYMERASE CHAIN REACTION (PCR)

Amplification of the *stx1*, *stx2f*, *stx2*, *stx2c*, *stx2e*, *eaeA*, *stx2d* and *hlyA* gene was performed using primer pairs (Wang et al., 2002).

PCR was performed as follows in a 0.2 ml thin walled tube.

Components	Quantity
Nuclease free water	15.75 µl
Genomic DNA (0.2 µl/ µl)	2.25 µl
Forward Primer (10 µM)	0.75 µl
Reverse Primer (10 µM)	0.75 µl
10X Reaction Buffer	2.5 µl
dNTP Mix	2.25 µl
Taq DNA polymerase (1 µ/µl)	0.75 µl
Total Volume	25 µl

The amplification was carried out in thermal cycler by setting the following temperatures and time periods for each step. Initial denaturation of 94°C for 3 minutes followed by

30 cycles of denaturation at 94°C for 30 seconds, annealing at 55°C for 30 seconds and extension at 72°C for 1 minute. Final extension was carried out at 72°C for 5 minutes. 10 µl of PCR products on 1.6% agarose gel along with 1 kb ladder DNA as a standard and run at 100 V for 60 min. The bands were stained with ethidium bromide and visualized under UV-transilluminator. (Sawhney and Singh 2002)

III. RESULTS

The isolate pe74 showed metallic green sheath on Eosin Methylene blue agar (figure 1). On genomic DNA isolation, isolate pe74 showed prominent band on agarose gel electrophoresis (figure 2). On PCR amplification isolate pe74 showed no amplified product for *Stx1* however only primer dimmer were observed in sample (figure 3). PCR analysis of isolate pe74 showed positive to *stx2f* with a band close to 150 bp and a band between 100 and 200 bp for *stx2c* (figure 4).

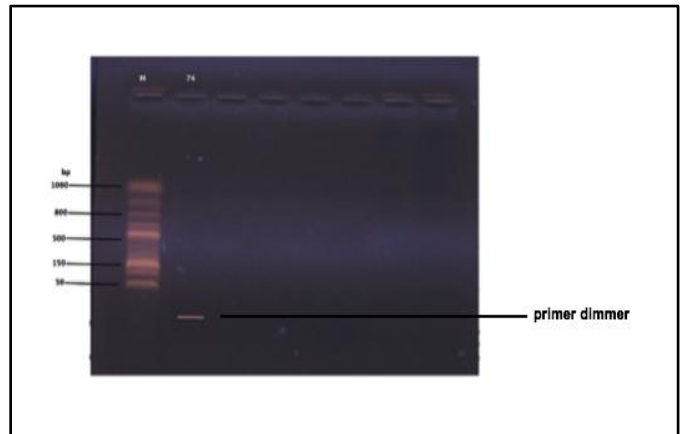


Figure 3: Agarose gel electrophoresis of PCR products of *stx1* gene from *E. coli* isolate pe74 Lane M: DNA molecular weight marker, Lane 1: pe74



Figure 1: Growth of isolate pe74 on Eosin Methylene Blue agar medium

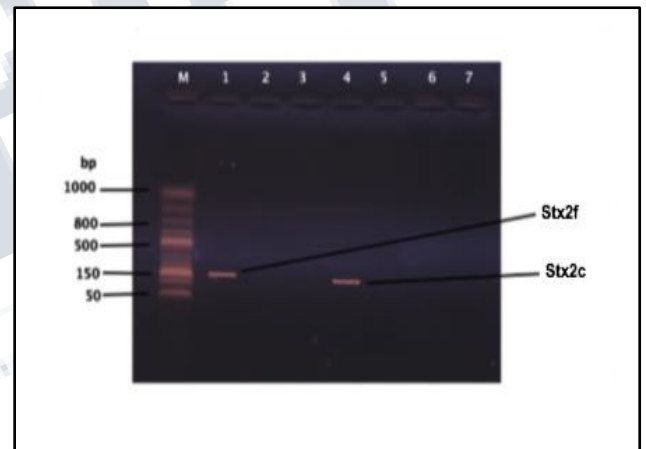


Figure 4: Agarose gel electrophoresis of PCR products of isolate pe74 Lane M: DNA molecular weight marker, Lane 1: *stx2f*, Lane 2: EHEC *hlyA*, Lane 3: *stx2*, Lane 4: *stx2c*, Lane 5: *eaeA*, Lane 6: *stx2b*, Lane 7: *stx2c*

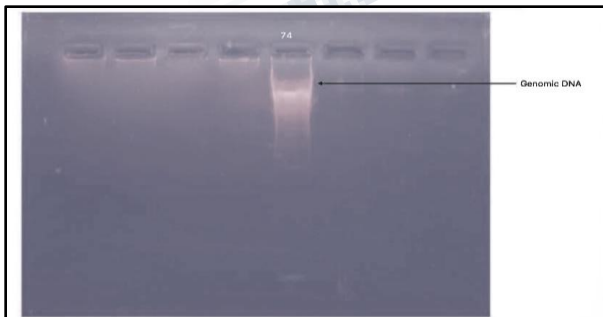


Figure 2: Agarose gel electrophoresis of genomic DNA from *E. coli* isolate pe74

IV. DISCUSSION

PCR analysis of isolate pe74 confirmed that *Stx2f* and *Stx2c* were detected. Among Shiga-toxin producing *Escherichia coli* *Stx2f* was recently reported to cause illness among human and difficult to detect immunologically (Skinner *et al.*, 2013).

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REFERENCES

1. Xiaodong, JianghongMeng, Patrick F McDermott, Sherry Ayers, Karen Blickenstaff, Thu-Thuy Tran, Jason Abbot, JieZheng and Saohuazhao. Presence and Characterization of Shiga Toxin-Producing *Escherichia coli* and other Potentially Diarrheagenic *E.coli* Strains in Retail Meats. *Applied Environment Microbiology*, 2010, 76(6): 1709-1717.
2. Riley L.W,Remis R.S, Helgerson S.D, McGee H.B, Wells J.G, Davis B.R, Hebert R.J, Olcott E.S, Johnson L.M, Hargrett N.T, Blake P.A, Cohen M.L, Hemorrhagic colitis associated with a rare *Escherichia coli* serotype, *Journal of Medicine*, 1983, 308: 681-685.
3. Sambrook J. and Russell Molecular cloning: A laboratory manual. Third edition vol 2, Cold Spring Laboratory Press, New York, 1989, pg-8.18-8.21, ISBN 087969-576-5.
4. Wang Gehua, Clifford G.Clark, and Frank G. Rodgers. Detection in *Escherichia coli* of the Genes Encoding the Major Virulence Factors, the Genes Defending the O157:H7 Serotype, and Components of the Type 2 Toxin Family by Multiplex PCR. *Journal of Clinical Microbiology*, 2002, 40(10): 3613-3619.
5. Sawhney S.K and Singh R. Introductory practical biochemistry, second reprint, Narosa publishing house, New Delhi, 2002, pg-415-416, ISBN 81-7319-302-9. Skinner C., Stephanie McMahan, ReuvenRasooly, John Mark Carter, and Xiaohua He. Purification and Characterization of Shiga Toxin 2f, an Immunologically Unrelated Subtype of Shiga Toxin 2. *PLoS One*, 2013, 8(3): e59760.